

09/068,528  
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(FILE 'HOME' ENTERED AT 08:22:18 ON 04 SEP 2003)

INDEX 'ADISCTI, ADISINSIGHT, ADISNEWS, AGRICOLA, ANABSTR, AQUASCI,  
BIOBUSINESS, BIOCOMMERCE, BIOSIS, BIOTECHABS, BIOTECHDS, BIOTECHNO, CABA,  
CANCERLIT, CAPLUS, CEABA-VTB, CEN, CIN, CONFSCI, CROPB, CROPU, DDFB,  
DDFU, DGENE, DRUGB, DRUGLAUNCH, DRUGMONOG2, ...' ENTERED AT 08:22:30 ON  
04 SEP 2003

SEA UDP-GLUCOSE

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35 FILE BIOBUSINESS  
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26 FILE CEABA-VTB  
1 FILE CEN  
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L1

FILE 'CAPLUS, BIOSIS, SCISEARCH, MEDLINE, EMBASE, TOXCENTER, PASCAL,  
CABA, BIOTECHNO, ESBIODBASE, LIFESCI, USPATFULL' ENTERED AT 08:23:38 ON 04  
SEP 2003

L2 7668 S L1 AND (SYNTHE? OR BIOSYNTHE?)  
L3 15 S L2 AND (GLUCOSE-1-PHOSPHATE URIDYLTRANSFERASE)  
L4 15 DUP REM L3 (0 DUPLICATES REMOVED)  
L5 364 S GDP-GLUCOSE OR GDP-SUGAR  
L6 248 S L5 AND (SYNTHE? OR BIOSYNTHE? OR MANUFACTU? OR METHOD OF MAK  
L7 8 S L6 AND GUANYLYLTRANSFERASE  
L8 8 DUP REM L7 (0 DUPLICATES REMOVED)

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COST IN U.S. DOLLARS

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TOTAL

multifunctional and did not have phosphomannose isomerase or phosphoglucose isomerase activity. Also, in contrast to the pig liver enzyme which uses mannose-1-P or glucose-1-P plus GTP to **synthesize** either GDP-mannose or **GDP-glucose**, the mycobacterial enzyme was specific for mannose-1-P as the sugar phosphate substrate. The enzyme was also relatively specific for GTP as the nucleoside triphosphate substrate. ITP was about 18% as effective as GTP, but ATP, CTP, and UTP were inactive. The activity of the enzyme was inhibited by **GDP-glucose** and glucose-1-P, although neither was a substrate for this enzyme. The pH optimum for the enzyme was 8.0, and Mg<sup>2+</sup> was the best cation with optimum activity at about 5 mM. This enzyme is important for producing the activated form of mannose for formation of cell wall lipoarabinomannan and various mannose-containing glycolipids and polysaccharides.  
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L8 ANSWER 5 OF 8 MEDLINE on STN  
 ACCESSION NUMBER: 93352609 MEDLINE  
 DOCUMENT NUMBER: 93352609 PubMed ID: 7688733  
 TITLE: GDP-mannose pyrophosphorylase. Purification to homogeneity, properties, and utilization to prepare photoaffinity analogs.  
 AUTHOR: Szumilo T; Drake R R; York J L; Elbein A D  
 CORPORATE SOURCE: Department of Biochemistry and Molecular Biology, University of Arkansas for Medical Sciences, Little Rock 72205-7199.  
 CONTRACT NUMBER: DK-21800 (NIDDK)  
 SOURCE: JOURNAL OF BIOLOGICAL CHEMISTRY, (1993 Aug 25) 268 (24) 17943-50.  
 Journal code: 2985121R. ISSN: 0021-9258.  
 PUB. COUNTRY: United States  
 DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)  
 LANGUAGE: English  
 FILE SEGMENT: Priority Journals  
 ENTRY MONTH: 199309  
 ENTRY DATE: Entered STN: 19931001  
 Last Updated on STN: 19960129  
 Entered Medline: 19930916

*Source is not a microorganism*

AB Pig liver GDP-mannose pyrophosphorylase was purified 5,000-fold to apparent homogeneity using standard techniques. The native enzyme showed a single band on gels of about 450 kDa and two subunits of 43 and 37 kDa on sodium dodecyl sulfate-polyacrylamide gel electrophoresis. The 37-kDa (beta-) subunit had only methionine at its amino terminus and a surprisingly hydrophobic sequence: Met-Lys-Ala-Leu-Ile-Leu-Val-Gly-Gly-Tyr-Gly-Thr-Arg-Leu-Arg-Pro-Leu-Thr-Leu-Ser-Ile-Pro-Lys. The 43-kDa (alpha-) subunit was blocked at the amino terminus, but a 29-kDa CNBr fragment had the following sequence: Leu-Asp-Ala-His-Arg-His-Arg-Pro-His-Pro-Phe-Leu-Leu-. Substrate specificity studies done in the direction of formation of nucleoside triphosphate and sugar-1-P indicated that the enzyme was most effective with **GDP-glucose** as substrate (100%) followed by IDP-mannose (72%) and then GDP-mannose (61%). That GDP-mannose and **GDP-glucose** activities were indeed catalyzed by the same enzyme was indicated by the following. (i) Various studies indicated that the enzyme was homogeneous. (ii) A staining procedure for production of GTP stained the same single band on native gels when either GDP-mannose or **GDP-glucose** was the substrate. (iii). GDP-mannose inhibited the utilization of **GDP-glucose** by the enzyme, and vice versa. When 8-azido-[32P]GTP was incubated with native enzyme and exposed to UV light, both the 43-kDa and the 37-kDa subunits became labeled, although the 37-kDa subunit reacted more strongly. On the other hand, 8-azido-GDP-[32P]mannose only photolabeled the 43-kDa band. Most importantly, the purified enzyme can be utilized to produce 8-azido-[32P]GDP mannose or 8-azido-[32P]**GDP glucose**.

L8 ANSWER 6 OF 8 CAPLUS COPYRIGHT 2003 ACS on STN

ACCESSION NUMBER: 1968:416342 CAPLUS  
DOCUMENT NUMBER: 69:16342  
TITLE: The purification and properties of guanosine diphosphate glucose pyrophosphorylase of pea seedlings  
AUTHOR(S): Peaud-Lenoel, C.; Axelos, M.  
CORPORATE SOURCE: Lab. Photosyn., C.N.R.S., Gif-sur-Yvette, Fr.  
SOURCE: European Journal of Biochemistry (1968), 4(4), 561-7  
CODEN: EJBCAI; ISSN: 0014-2956  
DOCUMENT TYPE: Journal  
LANGUAGE: English

AB A 400-fold purification of **GDP glucose** pyrophosphorylase from pea seedlings concomitant with stabilization of enzyme activity was accomplished by (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> fractionation, followed by chromatog. on Sephadex G-200 and DEAE-cellulose columns. In this last step, the enzyme is eluted with 0.20M NaCl. The enzyme is specifically activated by Mn<sup>2+</sup>. The apparent K<sub>m</sub> value for glucose 1-phosphate (37.degree., pH 7.5) is about 3 .times. 10<sup>-4</sup>M. The **GDP-glucose biosynthesis** is reversible, with the equil. in favor of pyrophosphorolysis. The purified enzyme is not active with GTP plus mannose 1-phosphate or galactose 1-phosphate. No formation of nucleotide sugar is observed in the presence of glucose 1-phosphate plus ATP, ITP, dTTP, or CTP.

L8 ANSWER 7 OF 8 CAPLUS COPYRIGHT 2003 ACS on STN

ACCESSION NUMBER: 1967:451963 CAPLUS  
DOCUMENT NUMBER: 67:51963  
TITLE: Nucleoside diphosphate glucose pyrophosphorylases in mast cell tumors  
AUTHOR(S): Danishefsky, Isidore; Heritier-Watkins, O.  
CORPORATE SOURCE: New York Med. Coll., New York, NY, USA  
SOURCE: Biochimica et Biophysica Acta (1967), 139(2), 349-57  
CODEN: BBACAQ; ISSN: 0006-3002  
DOCUMENT TYPE: Journal  
LANGUAGE: English

AB Mast cell tumors were shown to contain UDP-glucose pyrophosphorylase (UTP:.alpha.-D-glucose-I-phosphate uridylyltransferase EC2.7.7.9) and **GDP-glucose** pyrophosphorylase (GTP:.alpha.-D-glucose-I-phosphate **guanylyltransferase**). The 2 activities were sepd. from each other by (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> fractionation and are thus distinct enzymes. The **GDP-glucose** pyrophosphorylase fraction was purified further on DEAE-cellulose and some of its properties were studied. The enzyme was shown to catalyze the **synthesis** of **GDP-glucose** from D-glucose-I-phosphate and GTP. Other nucleoside triphosphates did not yield nucleoside diphosphate glucose when incubated with this enzyme prepn. The enzyme also catalyzed, to a lesser degree, the formation of GDP-mannose from GTP and D-mannose I-phosphate. Mannose I-phosphate also had an inhibitory effect on the **synthesis** of **GDP-glucose**. Fibrosarcomas and umbilical cord do not contain any detectable amt. of **GDP-glucose** pyrophosphorylase although they do have UDP-glucose pyrophosphorylase activity. 32 references.

L8 ANSWER 8 OF 8 CAPLUS COPYRIGHT 2003 ACS on STN

ACCESSION NUMBER: 1968:435 CAPLUS  
DOCUMENT NUMBER: 68:435  
TITLE: **GDP-glucose** pyrophosphorylase from peas  
AUTHOR(S): Barber, George A.  
CORPORATE SOURCE: Univ. of Hawaii, Honolulu, HI, USA  
SOURCE: Methods in Enzymology (1966), 8, 266-8  
CODEN: MENZAU; ISSN: 0076-6879  
DOCUMENT TYPE: Journal

LANGUAGE:

English

AB **GDP-glucose** pyrophosphorylase from peas catalyzes the reaction  $\text{GTP} + \text{glucose 1-phosphate} \rightarrow \text{GDP-glucose} + \text{inorg. pyrophosphate}$ . An assay method is described. The principle of the assay is the estn. of GDP-D-glucose- $^{14}\text{C}$  formed by adding  $^{14}\text{C}$ -labeled .alpha.-D-glucose 1-phosphate to the reaction mixt. The sugar nucleotide is isolated from the mixt. by adsorption on charcoal. The prepn. of the enzyme from fresh peas is described. The peas were homogenized and homogenate passed through cheesecloth and centrifuged at 20,000 g. The supernatant was made 0.5M in  $\text{MnCl}_2$ . The ppt. was discarded. The ext. was pptd. at pH 7.0 by 30-40% satn. with  $(\text{NH}_4)_2\text{SO}_4$ . The enzyme prepn. catalyzes the **synthesis** of D-glucosyl nucleotides and D-glucosyls of ADP, GDP, CDP, UDP, dTDP, IDP, and dUDP. The enzyme cannot use .beta.-D-glucose 1-phosphate, .alpha.-D-galactose 1-phosphate, .beta.-D-galactose 1-phosphate, or .alpha.-D-xylose 1-phosphate. The formation of GDP-D-glucose was stimulated about 3-fold by 5mM  $\text{MnCl}_2$  and about 1.5-fold by 5mM  $\text{MgCl}_2$ . Exts. of etiolated pea, mung bean seedlings, leaves of spinach, buckwheat, mustard, or parsley catalyze the **synthesis** of GDP-D-glucose from GTP and .alpha.-D-glucose 1-phosphate.

L5 ANSWER 511 OF 512 CAPLUS COPYRIGHT 2003 ACS on STN

ACCESSION NUMBER: 1965:45275 CAPLUS

DOCUMENT NUMBER: 62:45275

ORIGINAL REFERENCE NO.: 62:8069a-d

TITLE: The activation of *Escherichia coli*  
ADP-glucose **pyrophosphorylase**

AUTHOR(S): Preiss, , Jack; Shen, Laura; Partridge, Marian

CORPORATE SOURCE: Univ. of California, Davis

SOURCE: Biochemical and Biophysical Research Communications  
(1965), 18(2), 180-5  
CODEN: BBRCA9; ISSN: 0006-291X

DOCUMENT TYPE: Journal

LANGUAGE: English

AB *E. coli* B was grown in a **synthetic**, N-limiting medium at 37.degree. with glucose. Harvested cells were suspended in 4 vols. of 0.05M Tris-HCl buffer (pH 7.9), contg. 0.005M reduced glutathione. The cells were disrupted in a French press at 20,000 psi., and then centrifuged at 30,000 g for 15 min., the supernatant being used to supply **pyrophosphorylase**. ADP-glucose **pyrophosphorylase** (I) was purified 44-fold by protamine sulfate pptn., (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> fractionation, and heat treatment at 65.degree.. Phosphoglucomutase, aldolase, and **UDP-glucose pyrophosphorylase** were found in the crude supernatant, but were not found after purification. Enzyme activity was detd. from the **synthesis** of radio-ATP from ADP-glucose and inorg. pyrophosphate-32P (32PPi). Nucleoside triphosphate was sepd. from 32PPi by the use of Norit. I was stimulated 50-fold by fructose 1,6-diphosphate (II), whereas with phosphoenolpyruvate (III) or glyceraldehyde 3-phosphate(IV) it was stimulated 20-fold. Detn. of the II-concn. in the reaction mixt. with aldolase, triose phosphate dehydrogenase, DPN, and arsenate showed that its concn. did not change during the formation of ADP-glucose from ATP and glucose 1-phosphate. Of the tested compds., II, III, and IV demonstrated the best activation. Fructose 6-phosphate and pyruvate, which activated I from *Arthrobacter*, did not activate I from *E. coli*. 2,3-Diphosphoglycerate, glucose 6-phosphate, glucose, fructose 1-phosphate, NaHCO<sub>3</sub>, and succinate did not activate the enzyme. II did not activate **UDP-glucose**, TDP-glucose, and GDP-mannose **pyrophosphorylases** from the 30,000 g supernatant. I from *Aerobacter aerogenes* was activated by II, III, and IV. I from *Rhodo-spirillum rubrum* or *Agrobacterium tumefaciens* was activated by fructose 6-phosphate, pyruvate, and ribose 5-phosphate. Therefore, on the basis of activation, 2 types of I were distinguished.

L5 ANSWER 502 OF 512 CAPLUS COPYRIGHT 2003 ACS on STN  
ACCESSION NUMBER: 1972:69790 CAPLUS  
DOCUMENT NUMBER: 76:69790  
TITLE: Activity of nucleoside diphosphate sugar  
**synthetase (pyrophosphorylases)** in  
cells of *Escherichia coli* and *Shigella*  
*flexneri*  
AUTHOR(S): Janczura, Ewa; Chojnacki, Tadeusz  
CORPORATE SOURCE: Zakl. Bakterirol., Panstw. Zakl. Hig., Warsaw, Pol.  
SOURCE: Medycyna Doswiadczalna i Mikrobiologia (1971), 23(4),  
297-302  
CODEN: MDMIAZ; ISSN: 0025-8601  
DOCUMENT TYPE: Journal  
LANGUAGE: Polish  
AB Activities of the title enzymes were detd. in cell-free exts. of *E. coli* HfrC and in 8 strains of *S. flexneri* using <sup>32</sup>P-labeled glucose 1-phosphate and the appropriate nucleoside 5'-triphosphates. In *S. flexneri* only TDP-glucose was **synthesized**; the amt. of the nucleoside diphosphate sugar was twice as high in avirulent as in virulent strains. In *E. coli* exts., ADP, CDP, GDP, TDP, and IDP (but not UDP) glucose were **synthesiz**

L4 ANSWER 13 OF 15 EMBASE COPYRIGHT 2003 ELSEVIER SCI. B.V. on STN

ACCESSION NUMBER: 75104635 EMBASE

DOCUMENT NUMBER: 1975104635

TITLE: [Biosynthesis of UDP glucose  
by rat liver microsomes].

BIOSYNTHESE DE L'UDPGLUCOSE PAR LES MICROSOMES  
DES HEPATOCYTES DE RAT.

AUTHOR: Berthillier G.; Got R.

CORPORATE SOURCE: No 66 Lab. Biochim. Membranes, ERCNRS, Villeurbanne, France

SOURCE: Biochimica et Biophysica Acta, (1974) 362/2 (390-402).

CODEN: BBACAQ

DOCUMENT TYPE: Journal

FILE SEGMENT: 029 Clinical Biochemistry

023 Nuclear Medicine

LANGUAGE: French

AB Evidence is presented to show that all enzymes and all intermediary metabolites of a UDPglucose **biosynthesis** pathway are present in the microsomal membranes of rat liver. Glucose 6 phosphate, glucose 1 phosphate and UDPglucose are characterized by chromatography. The properties of phosphoglucomutase and UTP : D **Glucose 1 phosphate uridyltransferase** are studied. The K(m) values of phosphoglucomutase at pH 7.2 and 42.degree.C were  $0.26 \times 10^{-3}$  mM for glucose 1,6 diphosphate and  $80 \times 10^{-3}$  mM for glucose 1 phosphate. The K(m) values of UTP : D glucose 1 phosphate uridyl uridyltransferase at pH 8.5 and 37.degree.C were  $220 \times 10^{-3}$  mM for UTP and  $166 \times 10^{-3}$  mM for glucose 1 phosphate. These values are compared to the given values for enzymes from different species, and to those found for soluble enzymes. The significance of this membranous pathway is discussed.



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L8 ANSWER 1 OF 8 USPATFULL on STN

ACCESSION NUMBER: 2003:79307 USPATFULL  
TITLE: Active-site engineering of nucleotidylyltransferases  
and general enzymatic methods for the **synthesis**  
of natural and "unnatural" UDP- and TDP-nucleotide  
sugars  
INVENTOR(S): Thorson, Jon, Madison, NY, UNITED STATES  
Nikilov, Dimitar B., New York, NY, UNITED STATES

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 2003055235	A1	20030320
APPLICATION INFO.:	US 2001-13542	A1	20011213 (10)

	NUMBER	DATE
PRIORITY INFORMATION:	US 2000-254927P	20001213 (60)
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	KENYON & KENYON, 1500 K STREET, N.W., SUITE 700, WASHINGTON, DC, 20005	
NUMBER OF CLAIMS:	54	
EXEMPLARY CLAIM:	1	
NUMBER OF DRAWINGS:	32 Drawing Page(s)	
LINE COUNT:	3332	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

AB The present invention provides mutant nucleotidylyl-transferases, such as E.sub.p, having altered substrate specificity; methods for their production; and methods of producing nucleotide sugars, which utilize these nucleotidylyl-transferases. The present invention also provides methods of **synthesizing** desired nucleotide sugars using natural and/or modified Ep or other nucleotidylyltransferases; and nucleotide sugars synthesized by the present methods. The present invention further provides new glycosyl phosphates, and **methods** for **making** them.

L8 ANSWER 2 OF 8 USPATFULL on STN

ACCESSION NUMBER: 2002:272847 USPATFULL  
TITLE: Glycoconjugate and sugar nucleotide **synthesis**  
using solid supports  
INVENTOR(S): Wang, Peng G., Troy, MI, UNITED STATES  
Chen, Xi, Norristown, PA, UNITED STATES

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 2002150968	A1	20021017
APPLICATION INFO.:	US 2001-757846	A1	20010110 (9)
DOCUMENT TYPE:	Utility		
FILE SEGMENT:	APPLICATION		
LEGAL REPRESENTATIVE:	Brinks Hofer Gilson & Lione, P.O. Box 10395, Chicago, IL, 60610		
NUMBER OF CLAIMS:	43		
EXEMPLARY CLAIM:	1		
NUMBER OF DRAWINGS:	22 Drawing Page(s)		
LINE COUNT:	2405		

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

AB This invention relates to methods and compositions for the in vitro production of glycoconjugates. In particular, a preferred production system is provided that comprises a solid support, at least one sugar nucleotide producing enzyme, at least one glycosyltransferase, at least one bioenergetic, and at least one acceptor. The sugar nucleotide producing enzyme(s) is preferably immobilized on the solid support. The

glycosyltransferase may be co-immobilized on the solid support with the sugar nucleotide producing enzyme(s), or may be provided to the solid support in solution.

L8 ANSWER 3 OF 8 USPATFULL on STN

ACCESSION NUMBER: 2002:243134 USPATFULL  
TITLE: Glycoconjugate **synthesis** using a  
pathway-engineered organism  
INVENTOR(S): Wang, Peng George, Troy, MI, UNITED STATES  
Chen, Xi, Norristown, PA, UNITED STATES  
Liu, Ziyi, Detroit, MI, UNITED STATES  
Zhang, Wei, Detroit, MI, UNITED STATES

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 2002132320	A1	20020919
APPLICATION INFO.:	US 2001-758525	A1	20010110 (9)
DOCUMENT TYPE:	Utility		
FILE SEGMENT:	APPLICATION		
LEGAL REPRESENTATIVE:	BRINKS HOFER GILSON & LIONE, P.O. BOX 10395, CHICAGO, IL, 60610		
NUMBER OF CLAIMS:	51		
EXEMPLARY CLAIM:	1		
NUMBER OF DRAWINGS:	22 Drawing Page(s)		
LINE COUNT:	2558		

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

AB This invention relates to methods and compositions for the production of glycoconjugates. In particular, organisms are provided with at least one heterologous gene encoding an enzyme for regenerating a sugar nucleotide along with at least one glycosyltransferase. Such organisms are useful for the large-scale **synthesis** of glycoconjugates.

L8 ANSWER 4 OF 8 MEDLINE on STN

ACCESSION NUMBER: 1999145457 MEDLINE  
DOCUMENT NUMBER: 99145457 PubMed ID: 9989944  
TITLE: Purification and properties of mycobacterial GDP-mannose pyrophosphorylase.  
AUTHOR: Ning B; Elbein A D  
CORPORATE SOURCE: Department of Biochemistry and Molecular Biology,  
University of Arkansas for Medical Sciences, Little Rock,  
Arkansas, 72205, USA.  
CONTRACT NUMBER: R03-AI43292 (NIAID)  
SOURCE: ARCHIVES OF BIOCHEMISTRY AND BIOPHYSICS, (1999 Feb 15) 362  
(2) 339-45.  
Journal code: 0372430. ISSN: 0003-9861.  
PUB. COUNTRY: United States  
DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)  
LANGUAGE: English  
FILE SEGMENT: Priority Journals  
ENTRY MONTH: 199903  
ENTRY DATE: Entered STN: 19990324  
Last Updated on STN: 19990324  
Entered Medline: 19990311

AB The enzyme that catalyzes the formation of GDP-d-mannose from GTP and alpha-d-mannose-1-P was purified about 2300-fold to near homogeneity from the soluble fraction of Mycobacterium smegmatis. At the final stage of purification, a major protein band of 37 kDa was observed and this band was specifically labeled, and in a concentration-dependent manner, by the photoaffinity probe 8-N3-GDP[32P]-d-mannose. The purified enzyme was stable for several months when kept in the frozen state. The 37-kDa band was subjected to protein sequencing and one peptide sequence of 25 amino acids showed over 80% identity to GDP-mannose pyrophosphorylases of pig liver and Saccharomyces cerevisiae. In contrast to some other bacterial GDP-mannose pyrophosphorylases, the mycobacterial enzyme was not

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